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## Research Paper

One of the immune activation profiles observed in HIV-1-infected adults with suppressed viremia is linked to metabolic syndrome: The ACTIVIH study<sup>☆</sup>

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## ABSTRACT

Immune activation in HIV-1-infected individuals is reduced under antiretroviral therapies, but persists, resulting in various morbidities. To better characterize this phenomenon, using a panel of 68 soluble and cell surface markers, we measured the level of activation in circulating CD4+ and CD8+ T cells, B cells, monocytes, NK cells, polynuclear and endothelial cells as well as of inflammation and fibrinolysis in 120 virologic responders over 45 years of age. As compared with age- and sex-matched uninfected individuals, we observed a persistence of activation in all the cell subpopulations analyzed, together with marks of inflammation and fibrinolysis. Two independent hierarchical clustering analyses allowed us to identify five clusters of markers that varied concurrently, and five patient groups, each with the same activation profile. The five groups of patients could be characterized by a marker of CD4+ T cell, CD8+ T cell, NK cell, monocyte activation or of inflammation, respectively. One of these profiles was strongly associated with marks of metabolic syndrome, particularly with hyperinsulinemia (OR 12.17 [95% CI 1.79–82.86],  $p = 0.011$ ). In conclusion, our study unveils biomarkers linked to metabolic syndrome that could be tested as predictive markers, and opens the way to new therapeutic approaches tailored to each patient group.

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## 1. Introduction

Today, about 90% of treated persons living with HIV are aviremic (Delaugerre et al., 2015). Yet, 6 to 24% of these virologic responders do not restore their CD4 count as they should, exposing them to the risk of immune deficiency-linked morbidities (Corbeau and Reynes, 2011). The main reason for that is the persistence of a state of hyperactivity of the immune system in these patients (Corbeau and Reynes, 2011).

This residual immune activation also fuels the so-called non-AIDS-linked morbidities such as atherothrombosis, osteoporosis, metabolic syndrome, neurocognitive disorders, liver steatosis, kidney failure, frailty, and some types of cancer (Hunt, 2012). These comorbidities are now the major cause of morbi-mortality in treated patients (Lau et al., 2007).

HIV infection is characterized by the high level of immune activation induced in the human organism. Various causes may combine to provoke this immune hyperactivity (Younas et al., 2015). A first cause is the presence of the virus itself, detected by the immune system as a xenoantigen and a danger. For example, gp120 has recently been shown to activate monocytes and macrophages via TLR4 (Del Corno et al., 2016). A second cause is the infection process that triggers a DNA detection system resulting in inflammatory cytokine production (Doitsh

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et al., 2014). A third cause is the virus-induced deterioration of the gut barrier that opens the way for intestinal microbes to enter the human organism (Zevin et al., 2016). A fourth cause is the microbes that co-infect the organism (Boulougoura and Sereti, 2016), as for instance hepatitis (Kovacs et al., 2008) or herpes viruses (Sheth et al., 2008). A fifth set of causes is linked to some effects of HIV on the immune system itself: CD4+ T lymphopenia (Zonios et al., 2008; Sereti et al., 2009), immunosenescence (Effros et al., 2003) and an imbalance in the CD4+ T cell subpopulations (Kanwar et al., 2010). Finally, metabolic dysregulation may also be a source of immune activation (Aounallah et al., 2016; Dagenais-Lussier et al., 2015; Palmer et al., 2016). For instance, lipid peroxidation may fuel monocyte activation (Zidar et al., 2015). Moreover, the oxidative stress in the course of HIV infection might increase inflammatory cytokines production (Chaudhri and Clark, 1989). Combined antiretroviral therapies (cART) reduce all the sources of immune activation without abolishing them (Palmer et al., 2008; Guadalupe et al., 2003; Gonzalez et al., 2009; Stone et al., 2005). Consequently, immune activation is only partly downregulated in aviremic patients under treatment.

Previous works have shed a partial light on this phenomenon. The adaptive and the innate immune systems, including inflammation, are both involved. T cell activation has been particularly studied. Many authors have shown the persistence of marks of CD4+ and CD8+ T cell activation (Hunt et al., 2003), exhaustion (Cockerham et al., 2014), and senescence (Stone et al., 2005) under cART. The same is true for NK cells (Lichtfuss et al., 2012; Mavilio et al., 2005). Likewise, B cells remain quiet in virologic responders, as testified by the presence of hyperglobulinemia (De Milioto et al., 2004). In the same type of patients, high peripheral blood levels of soluble CD14 (sCD14) and soluble CD163 (sCD163) have also been reported (Lichtfuss et al., 2012; Burdo et al., 2011; Rajasuriar et al., 2010; Wallet et al., 2010; Kamat et al., 2012), considered as signs of monocyte/macrophage hyperactivity. Even neutrophils present with marks of activation in HIV-infected subjects, including high cell surface expression of the program death ligand 1 PD-L1 (Bowers et al., 2014) and of the adhesion molecules CD11b and CD18, as well as low cell surface expression of CD62L (Elbim et al., 1994), and PD-L1 expression has been shown to remain elevated on ART (Bowers et al., 2014). Inflammation is also evident in HIV patients, and for instance the routine marker CRP and the soluble form of TNF $\alpha$  receptor II has been reported to be incompletely reduced under treatment (Brazille et al., 2003; Aukrust et al., 1999; French et al., 2009; Calmy et al., 2009; Guimaraes et al., 2008; Regidor et al., 2011; Baker et al., 2011; van Vonderen et al., 2009; Funderburg et al., 2010; Shikuma et al., 2011). Various soluble markers have been used to monitor the activation of endothelial cells in the course of HIV infection, and some of them have been reported to be still increased after 12 years of ART (Rhönsholt et al., 2013). Finally, high levels of the main marker of fibrinolysis D-dimer have been observed (Kuller et al., 2008; Rodger et al., 2009; Neuhaus et al., 2010) that are not normalized under cART (Neuhaus et al., 2010).

Many studies have explored different aspects of this chronic immune, endothelial and coagulation activation, but we still lack a global picture of this phenomenon. Also, whether all patients have the same profile of activation or if various profiles may be distinguished remains unknown. This question is of major importance if we want to elucidate the specific role played by the different causes of immune activation, and for instance if some causes will favour some types of immune activation and other causes other types. It is also of importance if we want to have a better insight into the links between immune activation and comorbidities. A global analysis of immune activation might help us to assess if some profiles of immune activation favour specifically some morbidities. It is with this aim in mind, that we carried out an extensive characterization of the immune, endothelial and coagulation activation in HIV patients aviremic under treatment. This study enabled us to define five different profiles of immune activation in virologic responders.

## 2. Materials and methods

### 2.1. Study design

We carried out a cross-sectional observational study at two University Hospitals in France (Montpellier and Nîmes). Eligible patients were HIV-1-infected adults (age >45 years) with pretherapeutic CD4 cell counts of 350 cells per  $\mu$ L or less, CD4 cell counts of 200 cells per  $\mu$ L or more in the last 6 months, and plasma viral load below 50 copies per mL for four time points each separated by 6 months while on a potent, stable combination antiretroviral regimen. Exclusion criteria were: pregnancy, breastfeeding, immunomodulatory therapy, anticancer chemotherapy, active hepatitis B or C virus infection, decompensated cirrhosis, and any non-infectious disease susceptible to impact on the immune system. The Ethics Committee of the University Hospital of Montpellier approved this study. All patients provided written informed consent.

### 2.2. Flow cytometry

Monoclonal antibodies conjugated to fluorescein isothiocyanate (FITC), phycoerythrin (PE), energy-coupled dye (ECD), PE-Cyanine5.5 (PC5.5), PE-Cyanine7 (PC7), allophycocyanine (APC), APC/Alexa700, or APC/Alexa750 were purchased from Beckman Coulter. The antibodies were used in the following combinations; CD57-FITC/CD279-PE/CD45RA-ECD/C2-PC5.5/CD27-PC7/CD8-APC/CD4-APC700/CD3-APC750, CD8-APC/CD4-APC700/CD3-APC750/CD38-PE/HLADR-PC7/CD20-FITC, CD3-APC750/CD16 APC/HLADR-PC7/CD56-PC5.5/CD69-PE/CD57-FITC, CD4-FITC/CD45RA-ECD/CD25 PC7/FOXP3-APC/CD127-APC750. Whole blood collected into EDTA tubes was stained within 1 hour for 10 min at room temperature in the dark with cocktail of antibodies and fixed using IntraPrep Permeabilization Reagent Kit (Beckman Coulter) according to manufacturer's protocol. Cells were run on a Navios flow cytometer and results were analyzed by using Kaluza software (Beckman Coulter). A minimum 20,000 lymphocytes were gated to analyze the subpopulations. Representative gating strategies and images of flow plots are shown in Supplementary Fig. 1. We controlled the inter-run variability with the same batch of FlowSet Pro Beads (Beckman Coulter). The targets were defined on the samples settings (voltages). During the study, no voltage adjustment has been necessary to keep the beads into their respective defined targets.

### 2.3. Metabolic and soluble immunologic markers in peripheral blood

ELISA was used to quantify soluble TNF receptor I (sTNFRI), sCD14 and sCD163 (Quantikine, R&D systems), as well as tissue Plasminogen Activator, soluble Thrombomodulin, and soluble Endothelial Protein C Receptor (Asserachrom, Stago) in plasma collected into EDTA Vacutainer tubes (Becton Dickinson) and frozen. CRP and immunoglobulins were measured by turbidimetry in serum collected into SST II Vacutainer tubes, and D-dimers in blood collected into CTAD Vacutainer tubes (Becton Dickinson). Blood triglyceride, high-density lipoprotein (HDL) cholesterol, and insulin levels were quantified in serum collected into SST II Vacutainer tubes (Becton Dickinson) after 8 hours of fasting.

### 2.4. Statistical analysis

We used Wilcoxon tests to compare patients and donors, and Spearman rank correlations to evaluate the link between biomarkers with *p*-values multiplicity-adjusted by the False Discovery Rate method. We sorted the dataset with two independent hierarchical clustering analyses using respectively correlation and Euclidean distance to measure proximities/distances between markers and patients. The heatmap was generated using the *heatmap* function of the software R. The classifications used Ward (on the squared distance) as a metric. The distance was the euclidean for the patients and 1-abs (correlation) for markers. We used ANOVA results corrected by False Discovery Rate for multiple

testing, to determine the frequency of activation markers significantly different for at least one group of patients with regards to the other ones. Logistic regressions were carried out in order to study the relationship between profiles of immune activation and metabolic syndrome. Univariate analyses were first performed, and an adjustment by the age was thereafter executed. To study the relationship between activation markers of various components of the immune, endothelial and coagulation systems, we performed spearman correlations, and a Principal Component Analysis (PCA) after standardization of our variables. PCA allowed us to analyze multiple correlated values simultaneously, and to represent it as a set of new orthogonal variables called principal components, linear combinations of the variables. Then, the analysis of the components was carried out in order to highlight the degree of correlation between the variables. Statistical analyses were carried out using SAS Enterprise Guide V4.3 (SAS Institute Inc.), and R V3.1.1 (The R Foundation for Statistical Computing). A *p*-value lower than 0.05 was considered statistically significant.

### 3. Funding source

University Hospitals of Nîmes and Montpellier. The funding sources were involved neither in the study design, in the collection, analysis and interpretation of data, in the writing of the report, nor in the decision to submit the article for publication.

## 4. Results

### 4.1. Study subjects

Between April 29 and September 17, 2014, we recruited 22 female and 98 male HIV virologic responders (Table 1). Ninety-five percent of them were Caucasians. For all patients included, mean CD4 cell count was 688 (SD 326) cells per  $\mu\text{L}$  with a mean CD4:CD8 ratio of 0.99 (SD 0.52). Mean duration of HIV infection was 17.2 (SD 7.4) years with a mean pretherapeutic CD4 cell count of 192 (SD 108) cells per  $\mu\text{L}$ . 29% were current smokers, and 19% former smokers. The percentages of patients presenting with hepatitis A virus (HAV), hepatitis B virus (HBV), hepatitis C virus (HCV), cytomegalovirus (CMV), or Epstein-Barr virus (EBV) infection were 72%, 42%, 5%, 91%, and 98% respectively. Sex- and age-matched HIV-uninfected and HIV-infected, viremic, untreated caucasians were enrolled as negative and positive controls, respectively.

### 4.2. Activation markers

We selected markers known to be modified in untreated HIV patients, and potentially in treated HIV patients (Younas et al., 2015). At the T lymphocyte surface, we chose the activation markers HLA-DR and CD38, as well as the inhibitory receptor Programmed cell Death 1 (PD-1 or CD279). CD45RA and CD27 were used to distinguish naïve

**Table 1**

Bioclinical and therapeutic characteristics of the study populations. ND, not determined; NA, not applicable; NRTI, nucleoside reverse transcriptase inhibitor; NNRTI, non-nucleoside reverse transcriptase inhibitor.

Characteristic		HIV –	HIV + treated	HIV + untreated
Number of individuals		20	120	10
Age	Mean ( $\pm$ SD)	54.6 ( $\pm$ 6.4)	56.5 ( $\pm$ 8.0)	52.8 ( $\pm$ 10.2)
	Median (min–max)	53.0 (46.0–69.0)	55.0 (45.0–83.0)	50.5 (36.0–68.0)
Male sex	N (%)	16 (80)	98 (82)	8 (80)
Caucasians	N (%)	20 (100)	114 (95)	10 (100)
% CD4+ T cell	Mean ( $\pm$ SD)	ND	34.7 ( $\pm$ 9.8)	29.6 ( $\pm$ 10.7)
	Median (min–max)	ND	34.0 (15.0–62.0)	28.0 (16.0–47.0)
CD4 count (/mm <sup>3</sup> )	Mean ( $\pm$ SD)	ND	688 ( $\pm$ 326)	574 ( $\pm$ 283)
	Median (min–max)	ND	593 (243–2044)	509 (166–1153)
CD4:CD8 ratio	Mean ( $\pm$ SD)	ND	0.99 ( $\pm$ 0.52)	0.65 ( $\pm$ 0.33)
	Median (min–max)	ND	0.90 (0.20–3.30)	0.60 (0.20–1.20)
Pretherapeutic nadir CD4 count (cells/ $\mu\text{L}$ )	Mean ( $\pm$ SD)	NA	192 ( $\pm$ 108)	515 ( $\pm$ 304)
	Median (min–max)	NA	187 (1–458)	442 (130–1153)
Pretherapeutic viremia (RNA copies/ml)	Mean ( $\pm$ SD)	NA	393,985 ( $\pm$ 1,314,356)	58,833 ( $\pm$ 55,700)
	Median (min–max)	NA	104,156 (60–12,000,000)	35,234 (1220–170,318)
Years of HIV infection	Mean ( $\pm$ SD)	NA	17.2 ( $\pm$ 7.4)	1.1 ( $\pm$ 1.6)
	Median (min–max)	NA	18.0 (3.6–30.4)	0.3 (0.1–4.2)
Duration of viral suppression (months)	Mean ( $\pm$ SD)	NA	102 ( $\pm$ 47)	NA
	Median (min–max)	NA	95 (25–217)	NA
NRTI	N (%)	NA	108 (90)	NA
NNRTI	N (%)	NA	46 (38)	NA
Protease inhibitor	N (%)	NA	61 (51)	NA
Integrase inhibitor	N (%)	NA	34 (28)	NA
HBs Ag +	N (%)	ND	3 (2)	0 (0)
Anti-HBs Ab +	N (%)	ND	53 (44)	6 (60)
Anti-HBc Ab +	N (%)	ND	50 (42)	2 (20)
HCV coinfection	N (%)	ND	6 (5)	0 (0)
CMV coinfection	N (%)	ND	109 (91)	9 (90)
EBV coinfection	N (%)	ND	118 (98)	10 (100)
HAV coinfection	N (%)	ND	87 (72)	7 (70)
Systolic blood pressure (mmHg)	Mean ( $\pm$ SD)	ND	128 ( $\pm$ 16)	124 ( $\pm$ 14)
	Median (min–max)	ND	127 (100–181)	128 (100–136)
Diastolic blood pressure (mmHg)	Mean ( $\pm$ SD)	ND	80 ( $\pm$ 11)	82 ( $\pm$ 10)
	Median (min–max)	ND	80 (60–120)	82 (60–94)
Waist circumference (cm)	Mean ( $\pm$ SD)	ND	91 ( $\pm$ 11)	89 ( $\pm$ 10)
	Median (min–max)	ND	91 (65–128)	85 (82–108)
Lipodystrophy	N (%)	ND	40 (33)	0 (0)
Insulinemia ( $\mu\text{U/L}$ )	Mean ( $\pm$ SD)	ND	11 ( $\pm$ 7)	7 ( $\pm$ 5)
	Median (min–max)	ND	9 (1–45)	7 (2–17)
Triglyceridemia (mM/L)	Mean ( $\pm$ SD)	ND	1.86 ( $\pm$ 1.75)	1.55 ( $\pm$ 1.59)
	Median (min–max)	ND	1.57 (0.58–17.60)	1.10 (0.64–5.70)
HDL serum level (mM/L)	Mean ( $\pm$ SD)	ND	1.45 ( $\pm$ 0.61)	1.25 ( $\pm$ 0.55)
	Median (min–max)	ND	1.33 (0.16–4.20)	1.17 (0.50–2.42)

and central memory from effector memory T cells. CD57 upregulation along with CD27 and/or CD28 loss were used to characterize T lymphocyte senescence. In addition to these markers we also measured CD38 overexpression to evaluate CD8<sup>+</sup> T cell activation (Tuaille et al., 2009). NK cell activation was monitored analyzing HLA-DR and CD69 expression, NK cell dysfunction CD56 loss, and NK cell senescence CD57 gain. B cell hyperactivity was assessed by quantifying IgG, IgA, and IgM in the serum. The plasma level of sCD14 and sCD163 were used as indicators of monocyte/macrophage activation, and that of sTNFRI and C-reactive protein (CRP) as indicators of inflammation. Endothelium activation was explored by looking for an increase in soluble Endothelial Protein C Receptor (sEPCR) and in tissue Plasminogen Activator (tPA), and for a decrease in soluble Thrombomodulin (sThrombomodulin) plasma levels. In a subgroup of patients, D-dimer plasma concentration was measured as well as CD64, CD62L, and PD-L1 polynuclear surface density.

#### 4.3. Immune activation in virologic responders is global

As compared with HIV-negative controls, we observed in patients aviremic under cART an increase in the percentages of activated, HLA-DR<sup>+</sup>, CD4<sup>+</sup> T cells ( $16 \pm 8$  and  $22 \pm 13$ ,  $p = 0.015$ ), of activated, HLA-DR<sup>+</sup> ( $45 \pm 15$ ,  $54 \pm 18$ ,  $p = 0.042$ ), and senescent, CD57<sup>+</sup>, CD8<sup>+</sup> T cells ( $18 \pm 18$ ,  $32 \pm 14$ ,  $p = 0.001$ ), and of activated, HLA-DR<sup>+</sup> ( $33 \pm 22$ ,  $52 \pm 22$ ,  $p = 0.002$ ), dysfunctional, CD56<sup>−</sup> ( $8 \pm 4$ ,

$11 \pm 7$ ,  $p = 0.030$ ), and senescent, CD57<sup>+</sup> ( $33 \pm 16$ ,  $44 \pm 18$ ,  $p = 0.012$ ) NK cells (Fig. 1). Yet, these percentages were lower than those observed in non-treated HIV-infected individuals. We also observed an increase in the plasma level of sCD163 ( $279 \pm 158$  ng/mL,  $467 \pm 282$  ng/mL,  $p = 0.033$ ), and sTNFRI ( $0.9 \pm 0.3$  ng/mL,  $1.2 \pm 0.5$  ng/mL,  $p = 0.012$ ), even though these levels were below those measured in treatment-naïve patients. CD64 polynuclear surface density was higher in treated patients than in healthy controls ( $1.0 \pm 0.3$  versus  $0.8 \pm 0.1$  arbitrary units,  $p = 0.030$ ). As compared with virologic responders ( $95 \pm 56$  ng/mL) sThrombomodulin concentration was higher in control subjects ( $142 \pm 29$  ng/mL,  $p = 0.008$ ), but lower in viremic patients. Finally, IgG, IgA, IgM, and D-dimer plasma concentrations in aviremic patients were lower than in viremic patients, and higher than in noninfected controls (data not shown). Thus, all the components of the innate and adaptive immune system we tested, as well as the endothelium and the coagulation, appeared globally activated in the HIV-1-infected individuals we analyzed, even though they had been aviremic for years.

#### 4.4. Links between markers within each component of the immune and endothelial systems

We looked for correlations between the various markers within each cell subpopulation. In CD4<sup>+</sup> T cells (Table 2), expression of the

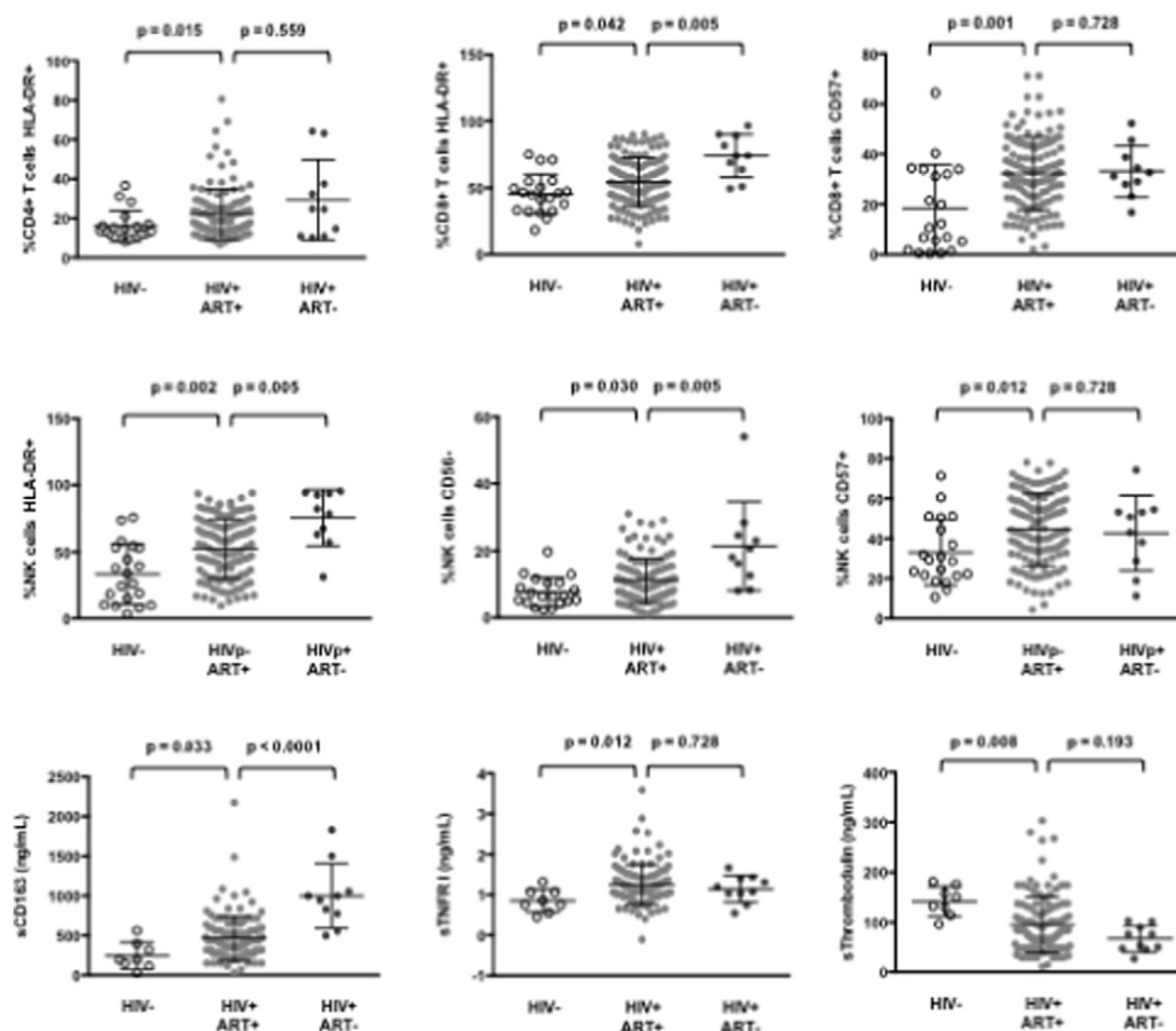


Fig. 1. Immune activation in virologic responders. Percentages of various cell populations and plasma levels of soluble markers in healthy donors (HIV-), treated (HIV+ ART+), and untreated (HIV+ ART-) HIV patients. Data are presented as mean values and 95% confidence intervals; p-values are shown.



activation marker HLA-DR was linked to that of the inhibitory receptor PD-1, and of markers of senescence (CD57 expression with and without loss of CD28 and/or CD27). In CD8+ T cells (Table 3), we found no correlation between HLA-DR and CD38 expression. Moreover, in these lymphocytes, HLA-DR expression, but not that of CD38, was linked to markers of senescence. Of note, in CD8+ T cells, PD-1 expression was linked neither to these two markers of activation, nor to markers of senescence. In NK cells (Table 4), the two activation markers HLA-DR and CD69 were linked together but not to the marker of senescence CD57, but this latter was linked to the marker of dysfunction CD56-. Of note, the levels of the monocyte/macrophage activation markers sCD14 and sCD163 were not linked together ( $r = 0.087$ ,  $p = 0.344$ ). By contrast, the plasma concentrations of IgG and IgA ( $r = 0.424$ ,  $p < 0.001$ ), and IgG and IgM ( $r = 0.352$ ,  $p < 0.001$ ) were linked together.

#### 4.5. Correlations between the various markers of the immune, endothelial, and coagulation activations

As we observed a hyperactivity in all the components we analyzed, we wondered whether the level of activity of some of these components were linked, either because their activation results from common causes and/or because they induce each other. To answer this question, we systematically looked for correlations between markers characterizing the activation of the various components of the immune, endothelial and coagulation systems (Table 5). As characteristic markers, we chose HLA-DR expression on CD4+ T cells, CD38 overexpression on CD8+ T cells, CD69 expression on NK cells, CD62L expression on polynuclear cells, sCD14, IgG, tPA, D-dimer and sTNFRI plasma levels for monocyte, B cell, endothelial, coagulation activation and inflammation, respectively. The only links we uncovered was between HLA-DR expression on CD4+ T cells and sCD14 level ( $r = 0.200$ ,  $p = 0.028$ ) on one hand, and sTNFRI and D-dimer levels ( $r = 0.215$ ,  $p = 0.011$ ) on the other hand.

In order to comprehensively analyze more globally correlations between the various activation markers and to obtain a visual representation, a PCA was computed, after standardization of the variables (Fig. 2). We decided to keep two principal components after performing an elbow test. These two components represented about 35% of the model's variance ( $22.70 + 11.96$ ). The results corroborated the conclusions above. For instance, in CD4+ T cells, we found a strong correlation between markers of senescence (CD57 expression with and without loss of CD28 and/or CD27), and of activation (HLA-DR) and PD-1. By contrast, D-dimer levels appeared not to be linked to CD4+ T cell and CD8+ T cell markers.

#### 4.6. Identification of different profiles of immune activation in patients aviremic under cART

To test the hypothesis of the existence of different profiles of immune activation in patients aviremic under treatment, we sorted the patients and the 68 markers of lymphocyte, NK cell, monocyte,

endothelium activation and inflammation by two independent hierarchical clustering analyses (Fig. 3). This clustering outlined the correlations, or the absence thereof, existing between some markers. Some of these correlations were expected. For instance the correlation between the percentages of CD8+ T cells positive for HLA-DR and for CD57 shown in Table 3 is illustrated by their proximity in the heatmap (8th and 10th markers from the right end). This clustering also unveiled the unexpected absence of links between other markers, such as between sCD14 and sCD163, as previously mentioned, situated far apart from each other in the heatmap. It also showed unexpected links between other markers. For instance, the marker of NK activation HLA-DR was linked to the marker of endothelium activation tPA ( $r = 0.261$ ,  $p = 0.004$ ). On the other hand, tPA appeared also linked to monocyte/macrophage activation as estimated by the level of sCD163 ( $r = 0.244$ ,  $p = 0.007$ ).

The hierarchical clustering analysis also identified five groups of patients, presenting with very different profiles of immune activation, and 2 outliers (Fig. 3). The levels of more than 80% of the markers were on average significantly different for at least one group of patients with regards to the other ones. The diversity of the activation profiles was evident when selected activation markers were analyzed (Fig. 4). As compared with the other groups, group 1 (T8 profile) was characterized by a high percentage of central memory (CD45RA-CD27+) CD8+ T cells ( $31 \pm 13$  versus  $23 \pm 10\%$ ,  $p = 0.0007$ ), group 2 (inflammation profile) by a high level of the inflammation marker sTNFRI ( $1.5 \pm 0.3$  versus  $1.2 \pm 0.5$  mg/L,  $p = 0.040$ ), and group 3 (NK profile) by a high number of activated, CD69+, NK cells ( $41 \pm 43$  versus  $31 \pm 60$  cells/ $\mu$ L,  $p = 0.033$ ). Group 4 (T4 profile) had the highest proportion of CD4+ T cells expressing CD38 ( $65 \pm 9\%$  versus  $55 \pm 12\%$ ,  $p < 0.001$ ), and group 5 (monocyte profile) the highest level of the monocyte activation marker sCD163 ( $671 \pm 520$  ng/mL versus  $446 \pm 279$  ng/mL,  $p = 0.009$ ). To simplify the identification of the five profiles of immune activation, we looked for a small number of markers able to characterize each patient group. Using only 8 biomarkers, we were able to classify the patients in the five groups with an error rate of 8.5%.

#### 4.7. Relationship between a profile of immune activation and metabolic syndrome

According to various studies, the frequency of metabolic syndrome has been reported as 14%–26% (Tebas, 2008) in HIV patients. This syndrome is characterized by the presence of at least 3 of the following components: hypertension, elevated waist circumference or waist-hip ratio, impaired glucose tolerance, and high triglyceride and/or low HDL cholesterol (Alberti et al., 2009). As the metabolic syndrome has been linked to immune activation (Esser et al., 2014), we looked for a relationship between this non-AIDS-linked morbidity and the immunological profiles we defined, by carrying out logistic regressions. Univariate analyses showed that, as compared with the other groups, patient group 2 presented with higher mean systolic and diastolic blood pressures, higher waist circumference, and lower HDL cholesterol

**Table 2**  
Correlations between CD4+ T cell markers of activation.

	% CD4+ T cells HLA-DR+	% CD4+ T cells PD-1+	% CD4+ T cells CD57+	% CD4+ T cells CD57+CD28-	% CD4+ T cells CD57+CD28-CD27-
% CD4+ T cells HLA-DR+		$r = 0.393$ $p < 10^{-4}$	$r = 0.585$ $p < 10^{-4}$	$r = 0.584$ $p < 10^{-4}$	$r = 0.542$ $p < 10^{-4}$
% CD4+ T cells PD-1+			$r = 0.375$ $p < 10^{-4}$	$r = 0.374$ $p < 10^{-4}$	$r = 0.326$ $p < 10^{-4}$
% CD4+ T cells CD57+				$r = 0.998$ $p < 10^{-4}$	$r = 0.976$ $p < 10^{-4}$
% CD4+ T cells CD57+CD28-					$r = 0.976$ $p < 10^{-4}$
% CD4+ T cells CD57+CD28-CD27-					

**Table 3**

Correlations between CD8+ T cell markers of activation.

	% CD8+ T cells HLA-DR+	% CD8+ T cells CD38+	% CD8+ T cells CD38hi	% CD8+ T cells HLA-DR+CD38+	% CD8+ T cells PD1+	% CD8+ T cells CD57+	% CD8+ T cells CD57+CD28–	% CD8+ T cells CD57+CD28–CD27–
% CD8+ T cells HLA-DR+				$r = 0.615$ $p < 10^{-4}$		$r = 0.503$ $p < 10^{-4}$	$r = 0.494$ $p < 10^{-4}$	$r = 0.462$ $p < 10^{-4}$
% CD8+ T cells CD38+			$r = 0.598$ $p < 10^{-4}$	$r = 0.443$ $p < 10^{-4}$				
% CD8+ T cells CD38hi				$r = 0.354$ $p < 10^{-4}$				
% CD8+ T cells HLA-DR+CD38+						$r = 0.506$ $p < 10^{-4}$	$r = 0.493$ $p < 10^{-4}$	$r = 0.436$ $p < 10^{-4}$
% CD8+ T cells PD-1+							$r = 0.996$ $p < 10^{-4}$	$r = 0.905$ $p < 10^{-4}$
% CD8+ T cells CD57+								$r = 0.907$ $p < 10^{-4}$
% CD8+ T cells CD57+CD28–								
% CD8+ T cells CD57+CD28–CD27–								

levels, but these differences did not reach significance. The frequency of hyperinsulinemia (insulinemia  $>24.9$  mIU/L; OR 12.17 [95% CI 1.79–82.86],  $p = 0.011$ , Fig. 5A), and of high ( $>2.85$  mM) triglyceridemia (OR 4.18 [95% CI 1.08–16.19],  $p = 0.038$ , Fig. 5B) was significantly elevated in group 2 as compared with the other groups. Moreover, the proportion of patients presenting with lipodystrophy was also higher in group 2 than in the other groups (OR 4.87 [95% CI 1.36–17.39],  $p = 0.015$ , Fig. 5C). Thus, the immune activation profile 2 was linked to major marks of one of non-AIDS comorbidities, the metabolic syndrome. Univariate analysis showed that, as compared with the other groups, patients in group 2 were slightly older (OR 1.08 [95% CI 1.01–1.16],  $p = 0.023$ ). A multivariate analysis, adjusted with age as a covariate, still highlighted the risk of lipodystrophy (OR 4.37 [95% CI 1.17–16.33],  $p = 0.028$ ) and a risk of hyperinsulinemia that was even higher (OR 17.06 [95% CI 2.14–135.60],  $p = 0.007$ ) in the group 2.

## 5. Discussion

In this study, we report a global evaluation of the state of activation of the immune, endothelial and coagulation systems in 120 treated patients aviremic for at least 2 years. A first conclusion is that these patients overall present with a generalized hyperactivity of these systems.

A second conclusion is that many markers of the activation of a cell subpopulation are not equivalent. Analyzing intra-subpopulation correlations, we noticed that HLA-DR expression and CD38 overexpression on CD8+ T cells were not correlated (Table 3). This is in line with the fact that these markers may not be coexpressed in vitro (Chadburn et al., 1992) and in vivo (Espinosa et al., 2013). It is also in agreement with the initial observation that CD38, but not HLA-DR, is a predictive marker of the disease progression (Giorgi et al., 1993). Clearly, the expression of these two markers is largely independent, probably driven by different mechanisms of activation, and leading to different consequences. Accordingly, CD8+ T cell senescence, as indicated by CD57 appearance and CD27 and/or CD28 loss, correlated with HLA-DR, but not

with CD38 expression (Table 3). Strikingly, PD-1 expression was linked to none of these two activation markers (Table 3). This emphasizes the notion that the percentage of CD8+ T cells positive for PD-1 is dependent on the stage of differentiation, PD-1 being mostly expressed at an early and intermediate stage of differentiation, rather than on the level of activation (Sauce et al., 2007). Indeed, in our study, the percentage of CD8+ T cells expressing PD-1 was very strongly (data not shown) correlated with the percentage of CD8+ T cells CD45RA-CD27+. Consequently, alternative markers need to be used to measure exhaustion, as for instance CD160 (Vigano et al., 2014). A striking observation is also that sCD14 and sCD163 are situated far apart in the biomarker clustering. The absence of correlation between these two monocyte/macrophage activation markers has been reported elsewhere (Patro et al., 2016). This is in line with the fact that sCD163, but not sCD14, has been correlated with HIV load, and may normalize under antiretroviral therapy (Castley et al., 2014). Likewise, sCD163, but not sCD14, has been linked to the level of anti-CMV IgG (Hodowanec et al., 2015) and to liver fibrosis (Mueller et al., 2015). Altogether these data emphasize the idea that these two markers likely do not reflect the same activation pathway.

Another important finding of our study is that the various pathways we analyzed are not often activated concurrently. In the individuals we examined, most of the components of the immune, endothelial and coagulation systems, as evaluated by selected markers, appear to be independently activated. This raised the interesting hypothesis that virologic responders might present with different profiles of activation.

The ascendant hierarchical classification enabled us to cluster activation markers that fluctuate in the same way. Apart from some expected correlations, this classification uncovered some unexpected ones. Indeed, to our knowledge, correlations between NK and endothelium activation had never been previously reported in HIV infection. The fact that activated endothelial cells may release fractalkine, a chemokine known to activate NK cells (Robinson et al., 2003) might be a mechanism warranting further study. Likewise, no links between monocyte/

**Table 4**

Correlations between NK cell markers of activation.

	% NK cells HLA-DR+	% NK cells CD69+	% NK cells HLA-DR+CD69+	% NK cells CD56–	% NK CD57+
% NK cells HLA-DR+		$r = -0.501$ $p < 10^{-4}$	$r = 0.395$ $p < 10^{-4}$		
% NK cells CD69+			$r = 0.270$ $p = 0.0029$		
% NK cells HLA-DR+CD69+					$r = -0.335$ $p = 0.0479$
% NK cells CD56–					$r = 0.270$ $p = 0.0002$
% NK cells CD57+					

**Table 5**

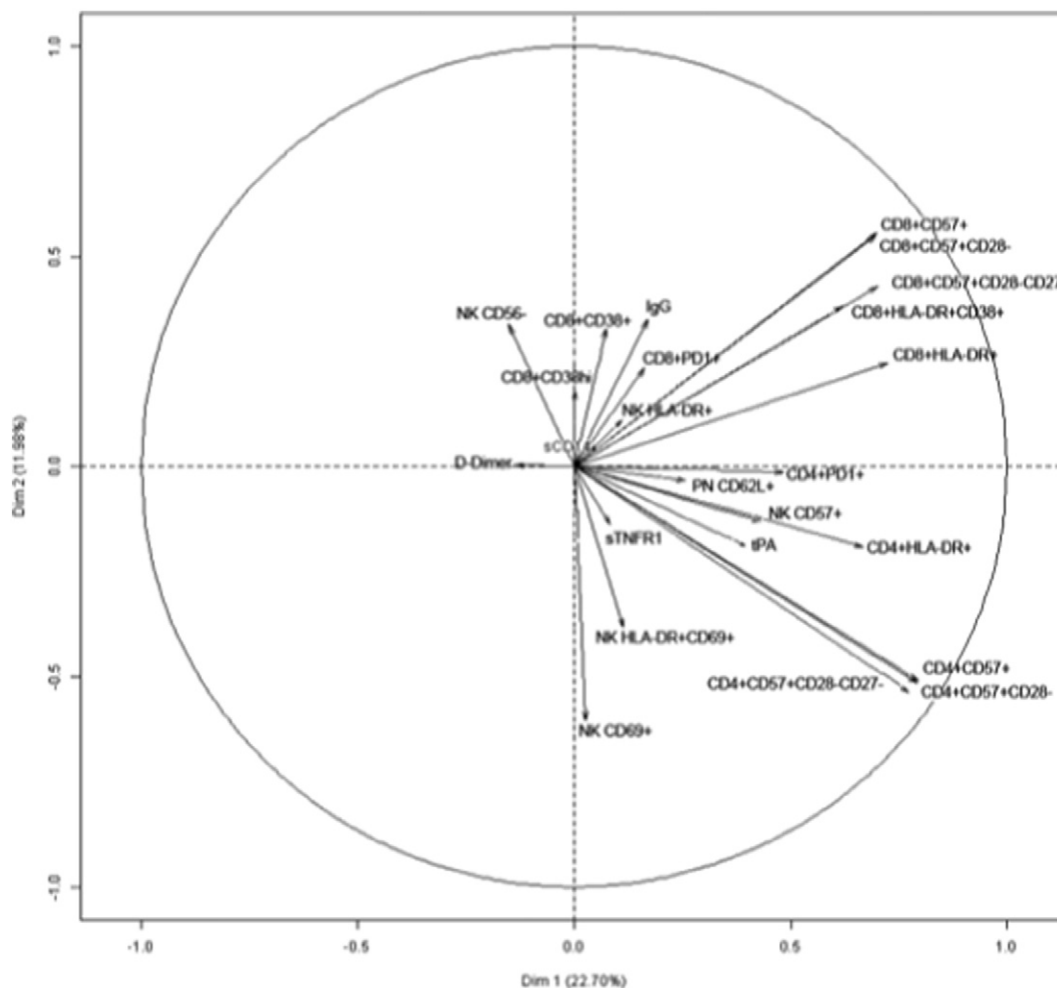
Correlations between markers of immune, endothelial and thrombolysis activation.

	% CD4+ T cells HLA-DR+	% CD8+ T cells CD38hi	% NK cells CD69+	CD62L polynuclear cell surface density	sCD14	IgG	sTNFR I	tPA	D-dimer
% CD4+ T cells HLA-DR+		–							
% CD8+ T cells CD38hi					$r = 0.200$				
% NK cells CD69+					$p =$				
CD62L polynuclear cell surface density					0.0284				
sCD14									
IgG									
sTNFR I							–		$r = 0.215$
									$p =$
									0.0107
tPA									
D-dimer									

macrophage and endothelium activation have been described so far in HIV patients. The fact that tPA has been reported to induce microglial activation via surface annexin II (Siao and Tsirka, 2002) might account for the correlation between tPA and sCD163 we uncovered. These examples highlight the interest of our global approach that may unveil

interactions between the components of the immune, endothelial and coagulation systems.

Via this global approach we were able to cluster the patients into five different immune activation profiles. The diversity of these profiles is illustrated by the fact that over 80% of the markers are variable among the



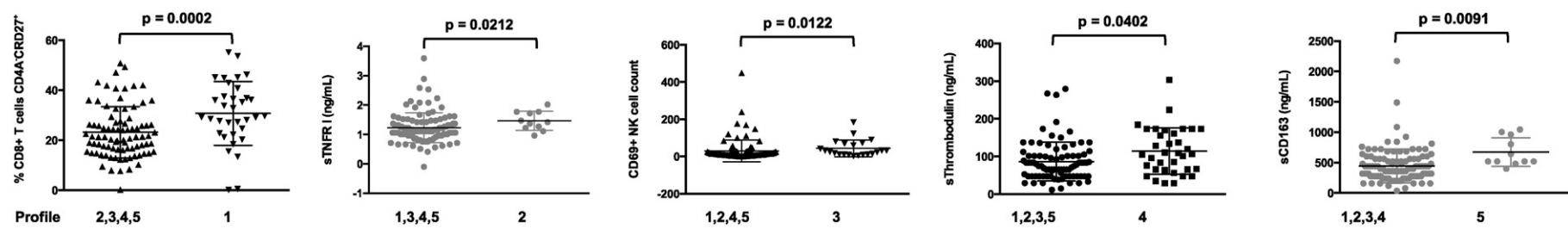
**Fig. 2.** Variables factor map resulting from Principal Component Analysis. The variables are represented by arrows. The elbow test was carried out in order to select the number of components to consider. This was done by plotting the components' eigenvalues according to their size and analyzing the point in the graph where the slope goes from "steep" to "flat" in order to keep only the components that are placed before the elbow, which were 2 in our case. The length of each of these arrows depends on the correlation of the variable with the component. Highly positively correlated variables are represented by arrows close to each other. Strongly negatively correlated variables are represented by arrows diametrically opposed.



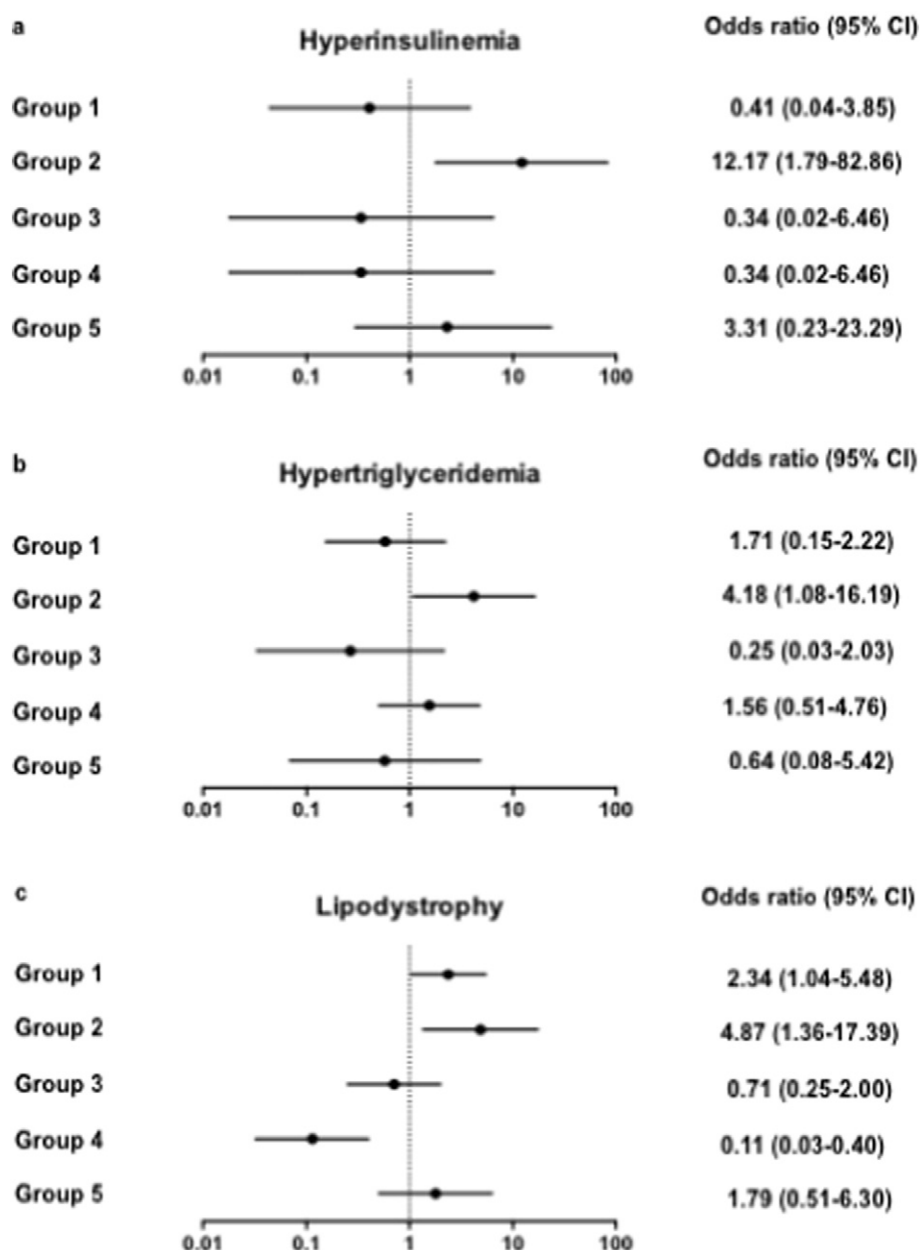
It is also possible that each activation profile fuels preferentially some comorbidities as compared with the other profiles. In line with this hypothesis, we observed a significant correlation between one of the patient group (group 2) and three hallmarks of the metabolic syndrome, hypertriglyceridemia, lipodystrophia and hyperinsulinemia. Of note, a characteristic of patients in group 2 is the high level of sTNFRI, a marker of TNF $\alpha$  production. Interestingly, TNF $\alpha$  has been reported to inhibit insulin signalling via the phosphorylation of the Insulin Receptor Substrate 1 (Hotamisligil et al., 1996; Yin et al., 1998; Aguirre et al., 2000), an effect that favours the development of metabolic syndrome (Alberti et al., 2009). Moreover, the plasma level of TNF $\alpha$  has been

From a more general point of view, it will be of interest to look for correlations between other comorbidities and the immune activation profiles uncovered by the present study. If we establish correlations between other comorbidities and some immune activation profiles, the simple signature of eight markers we have shown to be able to characterize each profile could be tested for its ability to predict some morbidities driven by immune/endothelial activation. Of practical interest, the identification of this signature requires a single flow cytometry tube. In the future, such a signature might enable to tailor prevention and early diagnosis of non-AIDS-linked morbidities to each patient. Unveiling links between some causes of immune activation and some immune activation profiles, and between some immune activation profiles and some comorbidities could also have therapeutic implications. Uncovering molecular mechanisms responsible for a causative link between a profile of immune activation and atherothrombosis, liver





**Fig. 4.** Characterization of the five different immune activation profiles. Differences in the levels of key activation markers between each group of patients and the other groups are represented. Data are presented as mean values and 95% confidence intervals; *p*-values are shown.



**Fig. 5.** Link between immune activation profile 2 and marks of metabolic syndrome. Odd ratios relating each profile of immune activation to risk of hyperinsulinemia (a), hypertriglyceridemia (b), and lipodystrophy (c). Data are presented as OR and 95% confidence intervals.

steatosis, osteoporosis or neurocognitive impairment could provide us with new therapeutic targets to prevent and attenuate these chronic diseases. Moreover, uncovering the causes (HIV and non-HIV infection, microbial translocation, metabolic dysregulation, CD4+ T cell loss, immunosenescence, and Treg dysfunction) fueling the immune activation profile of each patient could open the way to an etiologic treatment.

As these pathogenic pathways are probably not specific for HIV infection but relevant to any situation of chronic immune activation, including aging, and given the frequency of these comorbidities, these observations are of general interest. It is for instance striking that HIV infection and the autoimmune disease rheumatoid arthritis share many common comorbidities, including bone loss, atherosclerosis, metabolic syndrome, and frailty (Weyand et al., 2014). Thus, our approach could improve our knowledge on the pathogenic consequences of chronic immune activation, on the pathophysiology of the major chronic diseases it fuels, as well as on the prevention of these diseases, their early diagnosis and treatment.

Supplementary data to this article can be found online at <http://dx.doi.org/10.1016/j.ebiom.2016.05.008>.

#### Conflicts of Interest

None.

#### Authors Contributions

Christina PSOMAS contributed to the conception and design of the study, the patients' enrollment, acquired, analyzed and interpreted data. Mehwish YOUNAS contributed to the conception and design of the flow cytometry study, acquired, analyzed and interpreted cell surface and soluble markers data. Renaud CEZAR, Pierre PORTALES, Edouard TUAILLON, Jean-François ELIAOU contributed to the conception and design of the flow cytometry study, acquired, analyzed and interpreted cell surface markers. Christelle REYNES, Robert SABATIER,

Grégory MARIN, Nicolas NAGOT contributed to the conception and design of the statistical study, acquired, analyzed and interpreted statistical data. Adeline GUIGUES acquired and analyzed soluble markers data. Corinne MERLE, Nadine ATOUI, Céline FERNANDEZ, Vincent LE MOING, Claudine BARBUAT, Albert SOTTO acquired, analyzed and interpreted clinical data. Jacques REYNES contributed to the conception and design of the study, analyzed and interpreted data. Pierre CORBEAU contributed to the conception and design of the study, analyzed and interpreted data, and wrote the first draft of the manuscript. All authors revised and approved the final version.

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